

A causal association of genetically predicted serum 25- hydroxyvitamin D concentrations on the odds of tuberculosis disease: A Two-sample Mendelian Randomization Study

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Mercy W. Kimani

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Abstract

¹² Tuberculosis is a major cause of ill health and death. In 2018, an estimated 10 million people became ill and 1.2 million died from the disease. Vitamin D deficiency is also a major public health problem, affecting about a billion people globally. ¹² Studies have reported an association between vitamin D deficiency and an increased risk of tuberculosis. Some of the mechanisms underlying this association may include the role of vitamin D in both innate and adaptive immune functions. ¹¹ This study aimed to investigate whether there is a causal association between serum 25-hydroxyvitamin D (25(OH)D) levels and odds of tuberculosis disease in Asian and European populations using two-sample Mendelian randomization.

I used Mendelian randomization (MR), a technique that employs single nucleotide polymorphisms (genetic variants) as instrumental variables to estimate the causal effect of ²⁰ serum-25(OH)D (intervention) on the outcome of tuberculosis disease. I used genome-wide association studies (GWAS) summary statistics from the UK Biobank as the exposure data and from the International Tuberculosis Human Genetics Consortium (ITHGC) as the outcome data. Results from this study showed no evidence of a causal association of genetically predicted serum-25(OH)D concentrations on the odds of tuberculosis disease.



Declaration and Approval

I the undersigned declare that this dissertation is my original work and to the best of my knowledge, it has not been submitted in support of an award of a degree in any other university or institution of learning.

Signature

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In my capacity as a supervisor of the candidate's dissertation, I certify that this dissertation has my approval for submission.

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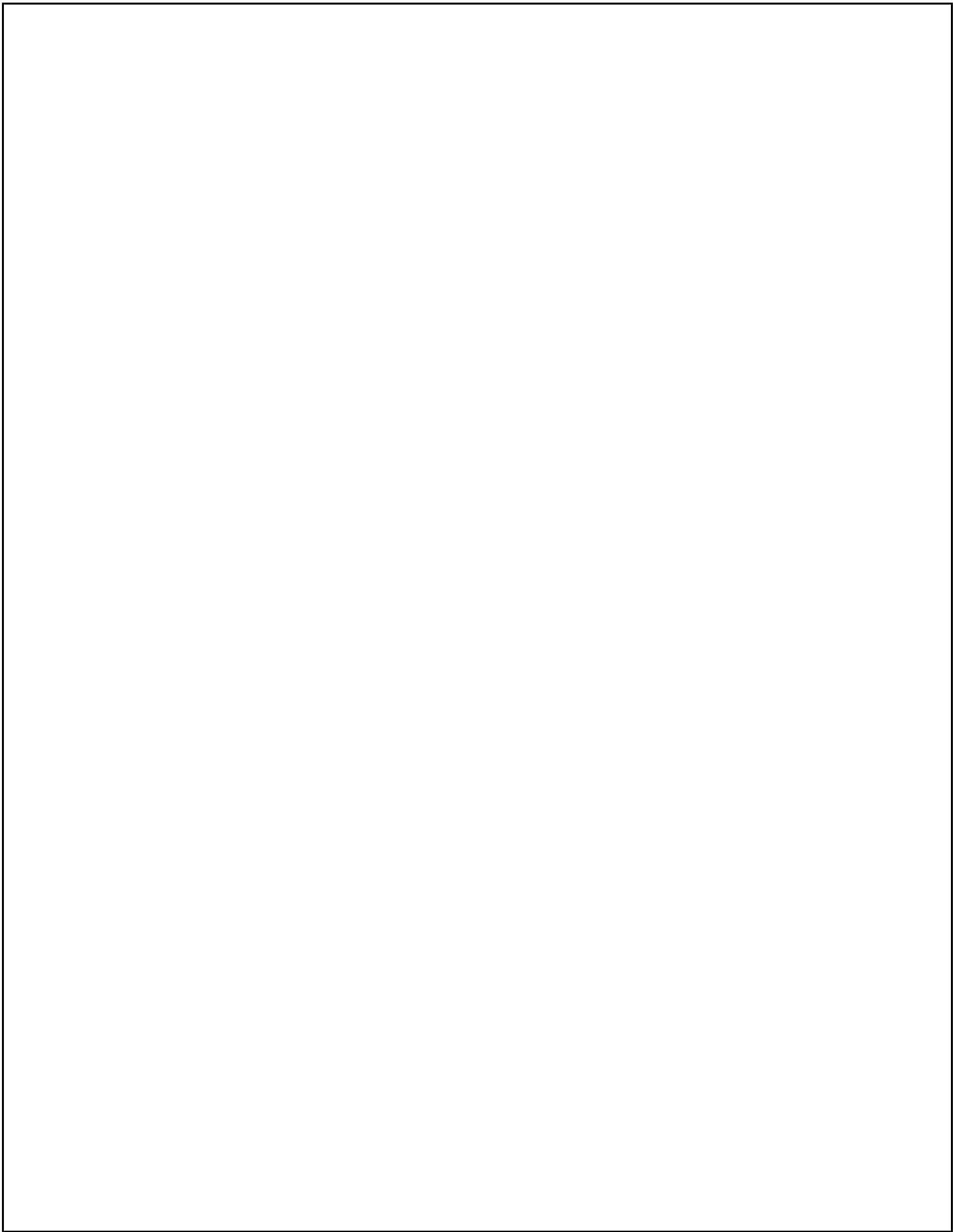
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Dedication

This project is dedicated to my parents and siblings.

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Mercy W. Kimani

Nairobi, 2020.

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1 Introduction

1.1 Background

Vitamin D deficiency and tuberculosis are both important global health concerns. Tuberculosis is a social and economic burden especially in South-East Asia, Africa, and the West Pacific, areas that account for 86-percent of cases (World Health Organization, 2020). An estimated 1.2 million people died from tuberculosis in 2019 making it one of the leading causes of death worldwide (World Health Organization, 2020). This is despite WHO reporting a two-percentage decline in the incidence of tuberculosis cases between 2018 and 2019 with an estimated ten million people becoming ill in 2019 (World Health Organization, 2020). Vitamin D deficiency affects almost a billion people globally with the greatest prevalence observed in the Middle East and Asia, especially among women (Holick, 2007; Natasja & Lips, 2011). A recent study has also reported that Africa is not an exception with approximately one in every five people having insufficient levels.4 Vitamin D deficiency is associated with the risk of both skeletal and non-skeletal diseases (Mogire et al., 2020). Some of the skeletal diseases include rickets in children, osteomalacia in adults and accelerated osteoporosis in older people (Bouillon et al., 2019). A causal association with multiple sclerosis has also been reported (Rhead et al., 2016). Many epidemiological studies provide evidence of an increased risk of tuberculosis (TB) in individuals with vitamin D deficiency (S. J. Huang et al., 2017; Nnoaham & Clarke, 2008; Venturini et al., 2014).

Genetic variants are used as instrumental variables to estimate whether there is a causal effect of an exposure (serum-25(OH)D levels) on an outcome (odds of tuberculosis), a technique called Mendelian Randomization (MR). In MR, genetic variants mimic the random assignment of treatments as in randomized controlled trials. Since genotypes are randomly allocated at meiosis, associations of the genetic variants with risk of tuberculosis will not be constrained by unaccounted for or unmeasured environmental risk factors (confounders). Also, since disease outcome cannot modify genotype, bias from reverse causality will be avoided (Sheehan, Didelez, Burton, & Tobin, 2008; Burgess & Thompson, 2015).

1.2 Statement of Problem

Many epidemiological studies have reported an association between vitamin D deficiency and an increased risk of tuberculosis disease. Randomized controlled trials (RCT) in children and individuals living with the human immunodeficiency virus (HIV) have recently

reported no ⁵ impact of vitamin D supplementation on the incidence of tuberculosis disease and infection. Further RCTs have been suggested and although they are the most robust means of evaluating potential interventions, they remain expensive and their capacity is limited. Mendelian ³⁴ randomization (MR) tests whether an exposure has a causal effect on an outcome by use of genetic variants as instrumental variables. Genetic variants are randomly assorted during meiosis, a process that mimics the random assignment of treatment in an RCT. This random allocation, as in a RCT, is independent of environmental factors meaning that exposure-outcome associations are unlikely to be a result of confounding by environmental exposures or reverse causation. This study seeks to explain whether serum vitamin D levels have a causal association with tuberculosis disease, which will also inform whether further RCTs of vitamin D for the prevention of tuberculosis are warranted.

1.3 Objectives

1.3.1 Overall objective

The general objective is to infer the causal effect of serum vitamin D levels on tuberculosis disease using two-sample Mendelian randomization.

1.3.2 Specific objectives

- I. ⁵ To determine the causal effect of vitamin D status on tuberculosis disease in a country-level analysis of different Asian and European countries.
- II. To estimate the overall causal effect of vitamin D status on tuberculosis disease in a meta-analysis of the Asian and European country-level results.

1.4 Significance of the Study

Tuberculosis is a major cause of ill health and death. There is, therefore, a need for intervention studies aimed at the prevention and treatment of tuberculosis. Findings from this study will help inform whether vitamin D deficiency has a causal effect on tuberculosis. Since vitamin D deficiency can be readily corrected for example by supplementing food products such as flour, the results from this study might help in the prevention of tuberculosis. Results will also inform whether the suggestion of further RCTs of vitamin D for the prevention of tuberculosis is warranted.

2 Chapter 2: Literature Review

2.1 An introduction to Tuberculosis

Tuberculosis (TB) is an infectious disease caused by bacteria of the *Mycobacterium tuberculosis* complex (MTC) (Castets, Boisvert, Grumbach, Brunel, & Rist 1968; Källenius et al. 1999; De Jong et al. 2009). Tuberculosis is thought to have spread to other parts of the world from Africa (Gutierrez et al. 2005). Ancient *Mycobacterium tuberculosis* (Mtb) strains, which originated from environmental mycobacteria, do not transmit in humans although they can still be isolated in immunocompromised individuals in some parts of East Africa (Supply et al. 2013; Delogu, Sali, & Fadda 2013). Increase in population density especially in urban areas has led to the emergence of a more virulent and highly transmissible strain called modern Mtb that is responsible for most cases of TB (Gagneux 2012; Wirth et al. 2008).

Mtb infection occurs when an individual inhales the aerosol droplets of tubercle bacilli. When these bacilli enter the alveoli of the lungs, they trigger innate immune responses (Urdahl, Shafiani, & Ernst 2011). Failure of the innate immune response to eliminate the pathogen leads to its multiplication inside the macrophage and later spread to other tissues and organs via the bloodstream and lymphatic fluid (Wolf et al. 2008; Balasubramanian et al. 1996). This triggers the cell-mediated immune functions that control the replication of the bacteria by forming a granuloma that encapsulates the bacilli (Ottenhoff & Kaufmann 2012). The granuloma later calcifies and resides in different organs and tissues in a dormant and metabolically inactive state called latent or dormant TB (Neyrolles et al. 2006; Hernandez-Pando et al. 2000; Feldman & Baggenstoss 1938). When cell-mediated immunity weakens, the bacteria replicates actively leading to active TB (Gengenbacher & Kaufmann 2012). Failure of the cell-mediated immune response to initially control the replication of the bacteria leads to the primary TB disease which is particularly seen in children (T. A. Thomas 2017). Ingestion of unpasteurised milk is also a route for transmission of zoonotic tuberculosis which is caused by *Mycobacterium bovis* (De la Rua-Domenech 2006). Tuberculosis manifests in different forms but the most common is pulmonary TB.

2.2 An overview of vitamin D

Vitamin D is a fat-soluble prohormone that exists in two forms, vitamin D3 (cholecalciferol), which is the animal form and vitamin D2 (ergocalciferol), which is the plant form. Vitamin D3 is primarily produced in the skin by ultraviolet B (UVB) irradiation on 7-

dehydrocholesterol, a precursor of cholesterol biosynthesis (Sylvia, DareV, Puneet, AdamJ, & Leilaj, 2010). Some vitamin D is also obtained from the diet through absorption in the intestine, which is essential for individuals not exposed to the sun, and for at-risk groups including growing children, pregnant women and the elderly (Zhu et al., 2012; Yun et al., 2017; Schilling, 2012). Vitamin D3 is converted to 25-hydroxyvitamin D (25(OH)D) in the liver, which is the main circulating form of vitamin D and which is used to measure serum-25(OH)D levels. 25(OH)D is later converted into 1,25-dihydroxyvitamin D (1,25(OH)2D) in the kidney, which is the active vitamin D metabolite (Sylvia et al., 2010). Serum vitamin D levels are subject to several factors including genetics, skin pigmentation, season, latitude, use of sunscreens, clothing style, nutrition and supplements (Natasja & Lips, 2011). Low serum vitamin D levels have also been reported among individuals suffering from chronic kidney disease (Franca Gois, Wolley, Ranganathan, & Seguro, 2018).

2.3 Association between vitamin D and TB

2.3.1 Vitamin D supplementation and TB trials

There are two randomized controlled trials (RCTs) testing the effect of vitamin D supplementation on the incidence of tuberculosis disease/infection that has been published so far. One of the RCTs enrolled 8,851 (4,418 in the supplementation group; 4,433 in the placebo group) children aged between 6 and 13 years old who were followed up for three years in Mongolia. The inclusion criteria were a negative result for Mtb infection at baseline according to the QuantiFERON-TB Gold In-Tube assay [QFT, Qiagen]. After a 3-year follow-up, results from this study showed that vitamin D supplementation (weekly oral dose of 14,000 IU of vitamin D3 supplements) did not affect the prevention of tuberculosis infection (adjusted risk ratio [aRR] 1.10; 95% confidence interval [CI] 0.87-1.38) or tuberculosis disease (aRR 0.87; 95% CI 0.49-1.55) (Ganmaa et al., 2020).

The second RCT study enrolled 4,000 (2,001 in the supplementation group; 1,999 in the placebo) adults above 18 years and living with the human immune-deficiency virus (HIV) who were initiating antiretroviral therapy (ART) at the time of randomization. The inclusion criteria were: a serum-25(OH)D concentration of less than 30 ng/ml as quantified with a commercial enzyme immunoassay (Immunodiagnosics; Boldon, UK); non-pregnant women; participants not enrolled in another trial; and intention to stay in Dar es -Salaam, Tanzania for at least a year. TB was diagnosed using sputum smears that were stained using the Ziehl-Nielsen technique to examine for acid-fast bacilli (AFB) and chest X-ray. The cartridge-based Xpert MTB/RIF (Xpert) assay (GeneXpert System; CA, USA) was used to test participants who were sputum AFB negative but still suspected for TB. Supplementation involved a weekly oral dose of 50000 IU of vitamin D3 supplements for the first month followed by a daily oral dose of 2000 IU of vitamin D3 supplements. After 1-year follow-up, results from this study also found no impact of Vitamin D supplementation on the incidence of pulmonary tuberculosis (hazard ratio [HR] 0.78, 95% CI

³¹ 0.54–1.13; $p=0.19$) or mortality (HR 1.04; 95% CI 0.85–1.25; $p=0.78$). (Sudfeld et al. 2020) They suggested that further RCTs might be warranted.

2.3.2 Observational studies of the ³⁰relationship between vitamin D and TB

The relationship between vitamin D and tuberculosis has been suggested since as early as the eighteenth century when fish oil was used to treat tuberculosis of the skin. (G. Rook 1988). In the 1940s, Solaria rooms that concentrate the sun's rays were used to treat tuberculosis (Mehta, 1939). Many epidemiological studies and meta-analyses also provide evidence supporting an association between vitamin D level⁹ and tuberculosis. The following are some examples of recent studies. Results from a nested¹⁷ case-control study, systematic review and individual-participant data⁹ meta-analyses of household contacts of TB patients in Peru reported that low serum vitamin D only predicts TB disease risk in a dose-dependent manner which strengthens a claim for a causal association. In this study, the meta³-analysis results revealed that vitamin D deficiency (<50 nmol/L) was associated with increased risk of progression to TB disease before (odds ratio [OR] 1.49; 95% CI 1.07–2.07; $p=0.02$), and after adjusting for age, gender, BMI, and HIV status (adjusted odds ratio [aOR] 1.48; 95% CI 1.04–2.10; $p=0.03$). Vitamin D insufficiency (50–75 nmol/L) was also associated with progression to TB disease after adjusting for age, gender, BMI, and HIV status (aOR 1.33; 95% CI 1.00–1.78; $p=0.05$) (Aibana et al. 2019). In their meta⁶-analysis, Huang and colleagues (S. J. Huang et al. 2017) found that vitamin D deficiency was associated with an increased risk of TB (OR 2.57; 95% CI 1.74–3.80; $P=0.00001$; $I^2=83$) with similar results being observed in a subgroup analysis of the Asian population (OR 2.62; 95% CI 1.63–4.23; $P=0.0001$; $I^2=71$). They also found that vitamin D deficiency was associated⁶ with an increased risk of developing active TB in latent TB infected (LTBI) subjects (OR 4.26; 95% CI 2.48–7.30; $P=0.00001$; $I^2=48\%$). Additionally, they reported that vitamin D deficiency was associated with an increased risk of tuberculin skin test (TST) conversion (OR 3.99; 95% CI 1.88–8.45; $P=0.0003$; $I^2=0$). In a meta-analysis of 3,998 immune-suppressed individuals, Zeng and colleagues (Zeng et al. 2015) reported an increased risk of active TB in individuals with serum 25(OH)D levels 12.5 nmol/l (pooled OR 4.556³; 95% CI 2.200–9.435; $I^2=11.9\%$; $P<0.001$) and within serum vitamin D levels between 13–25 nmol/L (pooled OR 3.797; 95% CI 1.935–7.405; $I^2=84.1\%$; $P<0.001$).

A cross-sectional study of household contacts in Mongolia reported that low levels of serum 25(OH)D were associated with latent tuberculosis infection (LTBI) (OR 0.92, 95% CI 0.88–0.97) after adjusting for age, sex, BMI, socio-economic, behavioural, and clinical characteristics²¹ (Gurjav et al. 2019). Additionally, a case-control study of prisoners in Brazil found that low serum vitamin D levels (<30 ng/ml) were associated with active TB in both univariate (OR 4.37; 95% CI 1.56–12.25) and multivariate analyses (aOR 3.77; 95% CI 1.04–13.64; after adjusting for drug use, previous incarceration and black race) (Maceda et al. 2018)²⁶. Pre-antiretroviral therapy (pre-ART) vitamin D deficiency was also associated with the risk of incident TB (adjusted hazard ratio [aHR] 3.66, 95%

CI 1.16–11.51) in a multinational case-cohort study of HIV-infected adults initiating ART (Tenforde et al., 2017). In a Korean case-control study, vitamin D deficiency (<20 ng/ml) was also associated with a higher odds of TB (OR 2.64; 95% CI 1.33–5.22) with severe vitamin D deficiency (<10 ng/ml) being associated with an even higher odds of TB (OR 11.75; 95% CI 6.53–21.15) (Hong et al., 2014). Venturini and colleagues also reported that vitamin D deficiency (<25 nmol/L) was correlated with TB infection (risk ratio [RR] 1.61; 95% CI:1.086–2.388; P=0.018) with a higher correlation reported in active-TB compared to latent TB and controls (RR 4.587; 95% CI: 1.190–9.608; P<0.0001), in a case-control study of children in the United Kingdom and Italy (Venturini et al., 2014). Moreover, a prospective study of 572 TB contacts in Spain found that reduced vitamin D levels (<20 ng/ml) were associated with risk of TB incidence (HR 0.88; 95%CI 0.80–0.97) (Arnedo-Pena et al., 2015).

2.3.3 Mechanisms underlying an association between Vitamin D and TB

Vitamin D has long been known for its role in the skeletal system. Studies on the extra-skeletal functions of vitamin D began in the late 20th century. These roles include lipid metabolism, cell proliferation and differentiation (Holick, 2003b, 1995, 2003a, DeLuca, 2004), insulin resistance (Chiu, Chu, Go, & Saad, 2004; Scragg, Sowers, & Bell, 2004), endocrine functions (DeLuca, 2004), inflammation (Gysemans et al., 2005; Haddad Kashani et al., 2018; Hossein-Nezhad et al., 2013) and immune functions (Cantorna, Zhu, Froicu, & Wittke, 2004; Yang, Smith, Prah, Luo, & DeLuca, 1993; Manolagas, Provvedini, & Tsoukas, 1985). The functions of vitamin D in the immune system span across both innate and adaptive host immune responses. 1,25-dihydroxyvitamin D induces production of the cathelicidin antimicrobial peptide (LL-37), in lymphocytes, neutrophils, human alveolar macrophages, and epithelial cells (Rivas-Santiago et al., 2008; Ashenafi et al., 2018). In vitro studies have shown that calcitriol (the active form of vitamin D, also known as 1,25-dihydroxy vitamin D), in these cells up-regulates the expression of LL-37 by the human cationic antimicrobial protein-18 (Hcap-18) gene (Martineau et al., 2007; Liu, Stenger, Tang, & Modlin, 2007; Rekha et al., 2015; Liu et al., 2006). LL-37 directly kills mycobacteria by osmotic lysis (Choi, Chow, & Mookherjee, 2012), and also induces autophagy, increases chemotaxis in macrophages, and regulates the production of chemokines and the secretion of cytokines (Skodric-Trifunovic et al., 2014). In vitro studies show that after infection, Toll-like receptors (TLR) are activated in human macrophages and monocytes (Liu et al., 2009). This increases the expression of vitamin D receptor and vitamin D-1-hydroxylase genes and therefore induces the antimicrobial peptide, cathelicidin which kills intracellular mycobacteria. When calcitriol binds to a membrane VDR on a Mtb infected macrophage, it induces a phagocyte superoxide burst which is mediated by a nicotinamide adenine dinucleotide phosphate (NADPH)-dependent phagocyte oxidase (Sly, Lopez, Nauseef, & Reiner, 2001; G. A. Rook et al., 1986). It also reduces the viability of MTB by enhancing phagolysosomal fusion in a phosphatidylinositol-3-kinases (PI3K)-dependent manner (Liu et al., 2009). Moreover, binding of 1,25-dihydroxyvitamin

D to the nuclear VDR upregulates innate functions such as induction of nitric oxide (NO) synthase and hence increased production of the toxic NO (Rockett et al., 1998).

2.4 GWAS studies

2.4.1 Genes associated with vitamin D status

Four genes associated with serum 25-hydroxyvitamin D (25(OH)D) concentrations have majorly been reported in different genome-wide association studies (GWAS). These genes are in the vitamin D synthesis and metabolism pathway (Figure 1). The group-specific component (GC) gene encodes the vitamin D binding protein (DBP), a protein that binds and transports vitamin D metabolites (Speeckaert, Huang, Delanghe, & Taes, 2006). The 7-dehydrocholesterol reductase (DHCR7) gene encodes an enzyme that converts 7-dehydrocholesterol to cholesterol using NADPH hence removing the substrate from the synthetic pathway of vitamin D₃. The nicotinamide adenine dinucleotide synthetase-1 (NADSYN1) gene encodes an enzyme involved in the last step in nicotinamide adenine dinucleotide (NAD) biosynthesis (Hara et al., 2003; Waterham & Wanders, 2000). The DHCR7 and NADSYN1 genes are together described as the DHCR7/NADSYN1 gene locus. The cytochrome P450 2R1 (CYP2R1) gene encodes microsomal vitamin D 25-hydroxylase, an enzyme that catalyses the hydroxylation of vitamin D₃ at carbon-25 to an active ligand for the vitamin D receptor (Cheng, Levine, Bell, Mangelsdorf, & Russell, 2004). The cytochrome P450 family 24 subfamily A member 1 (CYP24A1) gene encodes 24-hydroxylase, an enzyme that initiates the degradation of 1,25-dihydroxy vitamin D (1,25(OH)₂D) and 25(OH)D by catalysing the hydroxylation reactions at the side-chain (Jones, Prosser, & Kaufmann, 2012).

2.4.2 GWAS of Vitamin D

Genome-wide association study (GWAS) is a method of analysing genetic variations that are related to a phenotype by comparing genotype frequencies in different individuals in an observational study. Several GWAS of serum 25-hydroxyvitamin D (25(OH)D) concentrations has been done in populations of different ethnicities. GWAS of 417,580 individuals of European ancestry from the UK Biobank (UKB) identified 143 loci that were associated with 25(OH)D concentration. 18,864 single nucleotide polymorphisms (SNPs) were genome-wide significant ($P < 5 \times 10^{-8}$) with SNPs on the GC and CYP2R1 loci being the most associated ($P < 1.01 \times 10^{-400}$) (Revez et al., 2020). A different GWAS of 401,460 individuals of European ancestry (white British) from the UKB reported 69 loci that were associated with 25(OH)D concentration. Among these were the four genes on the vitamin D metabolism and synthesis pathway. The study also identified 138 genome-wide significant and conditionally independent SNPs ($P < 6.6 \times 10^{-9}$) (Manousaki et al., 2020). The SUNLIGHT (Study of Underlying Genetic Determinants of Vitamin D and Highly Related Traits) consortium GWAS of 79,366 European-ancestry individuals identified six

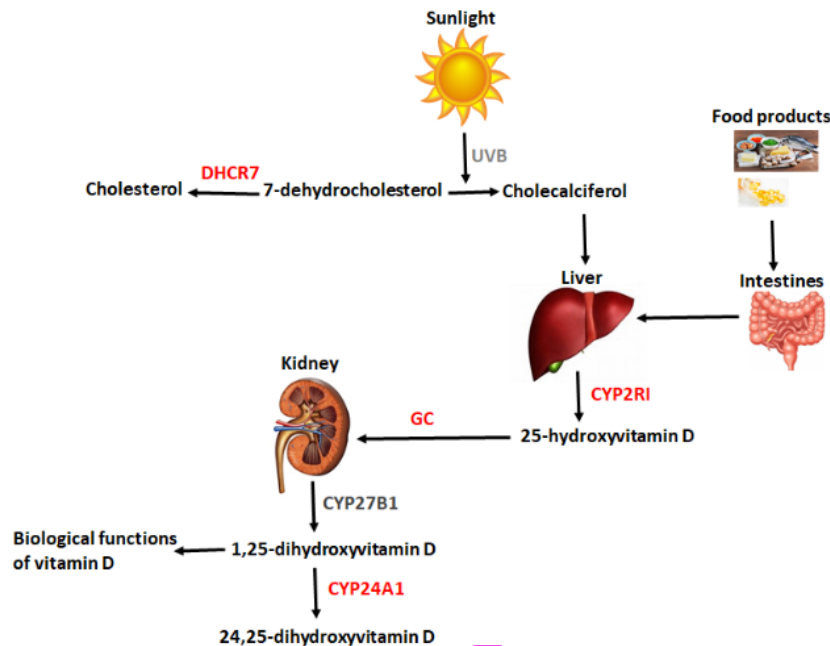


Figure 1. Genes associated with serum vitamin D levels ¹⁴

loci (GC, CYP2R1, DHCR7/NADSYN1, CYP24A1, Sec23 Homolog A, coat protein complex ¹⁴ II (COPII) component [SEC23A], Amidohydrolase domain containing 1 [AMDHD1]) to be associated with 25(OH)D (Jiang et al. 2018). Four of these loci had been reported in a previous GWAS of about 30,000 individuals of European descent by the same consortium (GC, CYP2R1, CYP24A1, DHCR7/NADSYN1) (T. J. Wang et al. 2010). An earlier GWAS of 4501 individuals of European ancestry had identified SNPs in the GC, DHCR7/NADSYN1 and CYP2R1 genes that were associated with 25(OH)D concentrations (Ahn et al. 2010). GWAS of 1,829 US women reported 32 SNPs located in or near ² GC and CYP2R1 loci that had genome-wide significant associations ($P < 5 \times 10^{-8}$) with 25(OH)D concentrations (O'Brien et al. 2018). GWAS of serum 25(OH)D on 3538 individuals from Asian (Indian) population in Punjabi reported six loci that were associated with 25(OH)D concentration with only one reaching the genome-wide significance (Forkhead Box A2 gene [FOXA2]: $p = 4.4710^{-9}$). The association between GC and CYP2R1 top variants ¹¹ with 25(OH)D concentrations in Europeans was also confirmed in the Punjabi sample (GC; $p = 0.007$ and CYP2R1; $p = 0.019$) (Sapkota et al. 2016).

2.4.3 Previous studies on vitamin D polymorphisms and TB

The genomic function of vitamin D is mediated by the vitamin D receptor (VDR). VDR ⁵ is a nuclear hormone receptor, present on monocytes, activated T and B lymphocytes, and many other cells and tissues, that binds 1,25-dihydroxyvitamin D (1,25(OH)₂D) (Y. Wang, Zhu, & DeLuca 2012). Once 1,25(OH)₂D binds to the nuclear VDR it reg-

ulates the transcription of vitamin D-responsive genes and also activates non-genomic signal transduction pathways in the cells (Bikle 2014, Gil, Plaza-Diaz, & Mesa 2018).

Currently, no study has tested the association between vitamin D polymorphisms on the GC, CYP2R1, CYP24A1 and DHCR7/NADSYN1 genes and the risk of TB. However, studies have found an association between VDR polymorphisms and TB. In a Taiwanese population, genotypes, AA of rs73126, GG of rs1544410 and A carriers of rs7041 were associated with susceptibility to TB (Lee et al. 2016). Three SNPs (rs11574143, rs11574079, and rs11168287) in the VDR gene were also associated with pulmonary tuberculosis in a Tibetan-Chinese population (Hu et al. 2016, Varahram et al., 2009). In a meta-analysis of 42 well-controlled studies, two variants (rs1544410 and rs731236) on the VDR gene were significantly associated with TB in the overall population and the south Asians in an ethnicity analysis (Xu & Shen 2019). In the subgroup analyses, the rs7975232 variant was associated with extra-pulmonary TB while the rs2228570 variant was associated with both pulmonary and extra-pulmonary TB.

2.5 Mendelian Randomization

2.5.1 Describing the technique

Mendelian randomization uses genetic variants as instrumental variables in observational data to determine if an exposure (risk factor) causes a disease outcome (D. C. Thomas & Conti 2004). Genetic variants proxy the differences in the risk factor. A genetic variant is a section of deoxyribonucleic acid (DNA), mostly a single nucleotide polymorphism (SNP) that differs between individuals. These genetic variants have to satisfy the instrumental variables assumptions which state that: (Greenland 2000) (Figure 2)

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- I. The genetic variant must be associated with the exposure.
- II. The genetic variant should not be associated with confounders of the exposure-outcome relationship.
- III. The genetic variant should only be associated with the outcome through the exposure.

In MR, instrumental variables mimic the random assignment of treatments as in an experimental study (Figure 3) (Nitsch et al. 2006). Although randomized controlled trials (RCTs) are the most robust means of evaluating interventions, they are expensive and their capacity is likely to remain limited. The advantage of using MR is that the method reduces bias due to confounding and reverse causation. This is because genetic variants are randomly assorted during meiosis and are therefore independent of environmental

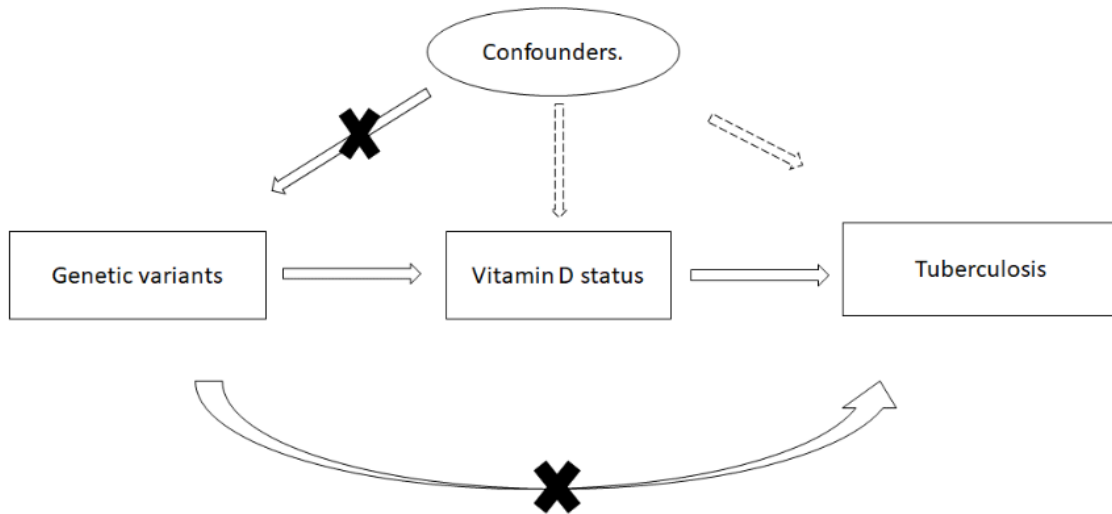


Figure 2. Assumptions of the genetic variants for Mendelian randomization

factors. MR has also been used to confirm the results of RCTs and to predict null results in RCTs costing upwards of USD 100 million (Labos, Brophy, Smith, Sniderman, & Thanassoulis, 2018; Yarmolinsky et al., 2018).

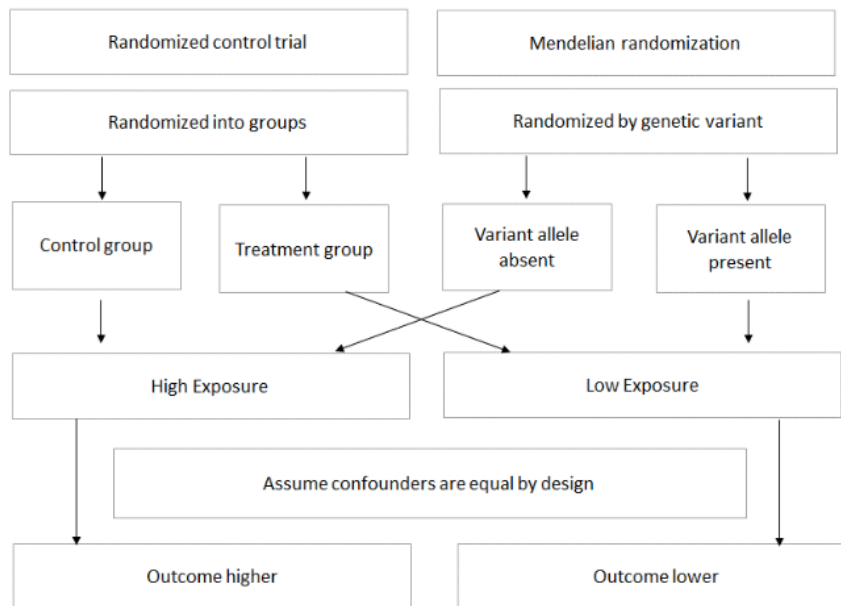


Figure 3. Comparison of Mendelian randomization and randomized controlled trial

Two sample-MR is an MR that uses summary statistics (that is beta-coefficients and standard errors) from two different, non-overlapping studies (GWAS of vitamin D and GWAS of TB in our study) to estimate whether a risk factor causally affects an outcome by estimating the IV-exposure and IV-outcome associations (Burgess et al., 2015). Summarized data are available from large consortia of genetic association studies where estimates

are obtained²⁷ from regression models of either the exposure or the outcome on a genetic variant. These data are often from large sample sizes, which increases the power of the analysis. The data can also be easily accessed and this allows reproducibility of the analysis and hence ensures transparency.

2.5.2 Previous vitamin D MR studies

Many Mendelian randomization (MR) studies between genetically predicted serum-25(OH)D⁵ levels and different phenotypes²⁹ have been conducted with some reporting a causal association. For example, vitamin D deficiency was found to have a causal relationship with increased all-cause mortality in two MR studies of 10,501 participants of European ancestry and 95,766 participants of Danish descent. These studies used genetic scores of SNPs on the DHCR7 and CYP2R1 loci to estimate the hazard ratio (HR) for mortality per 20 nmol/L decrease in genetically determined 25(OH)D levels. The HR estimates for the two genetic score in the European population study was 1.32 (95% CI: 0.84 to 2.20) and 1.35 (95%CI of 0.81 to 2.37) while that of the Danish population was 1.30 (95% CI: 1.051.61) (Aspelund et al., 2019; Afzal, Brøndum-Jacobsen, Bojesen, & Nordestgaard, 2014). The odds ratio (OR) of Alzheimer's disease (AD) per genetically-predicted one standard deviation increase in serum 25(OH)D was 0.86 (95% CI 0.78–0.94; $p = 0.002$) in a two-sample MR of 54,162 individuals of European ancestry (Larsson, Traylor, Markus, & Michaëlsson, 2018). A different MR of 54,162 individuals of European ancestry demonstrated that a one standard deviation decrease in natural log-transformed 25(OH)D increased the risk of AD (OR 1.25; 95% CI 1.03– 1.51; $p = 0.021$) (Mokry et al., 2016). Causal association between serum-25(OH)D and risk of ovarian cancer was reported in a two-sample MR study of 31,719 women of European ancestry with an OR of 1.17 (95% CI 0.76–1.78) per 20-nmol/L decrease in 25(OH)D levels (Ong et al., 2016). In a meta-analysis of ten studies of European and Asian populations (428,904 individuals), a 25 nmol/L increase in genetically determined 25(OH)D concentration was associated with a lower risk of diabetes (OR 0.86; 95% CI 0.77-0.97; $p = 0.01$) using a genetic score of DHCR7 and CYP2R1 synthesis SNPs (Lu et al., 2018). MR analyses of non-Hispanic whites (10,071 individuals) and Swedish population (12,097 participants) showed that increasing levels of 25(OH)D are associated with a decreased risk of multiple sclerosis (OR 0.79; 95% CI 0.64-0.99) and (OR 0.86; 95% CI 0.76–0.98) respectively. A meta-analysis of the two populations resulted in a combined OR of 0.85 (95% CI 0.76–0.94) (Rhead et al., 2016). One standard deviation decrease of genetically determined log-transformed 25(OH)D levels was also associated with an increased risk of multiple sclerosis (OR 2.02; 95% CI; 1.65–2.46; $p = 7.7210^{12}$) in an MR of 38,589 MS participants of European ancestry (Mokry et al., 2015). Genetically elevated levels of serum-25(OH)D was associated with a decreased risk of delirium (HR 0.74; 95% CI 0.62–0.87; $p = 0.0004$) in an MR of 313,121 participants of European descent from the UK biobank (Bowman et al., 2019).

3 Chapter 3: Methods

3.1 Data Sources for MR Analyses

I conducted a two-sample Mendelian randomization using summary statistics from non-overlapping Vitamin D and TB GWAS datasets. The Vitamin D GWAS datasets provided the exposure data while the TB GWAS datasets provided the outcome data. The flow diagram below shows the steps I took in conducting the MR (Figure 4).

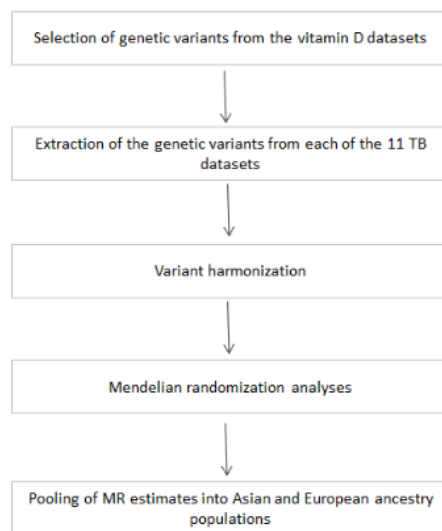


Figure 4. Flow diagram of analyses conducted.

3.1.1 Vitamin D GWAS Studies

I selected the instrumental variables for the European ancestry population, from the UK Biobank (UKB) vitamin D GWAS (Revez et al., 2020). The UKB is a population cohort of over 502,000 European individuals, aged between 40 and 69 years old who were recruited between 2006 and 2010 (Bycroft et al., 2018). Phenotypic, genotypic and clinical data were collected on these individuals. The North West Multicentre Research Ethics Service Committee approved the study and all participants signed informed consent. Individuals of European ancestry were identified using the 1000 Genome project reference cluster. Genome-wide genetic variation of the single nucleotide polymorphism (SNPs) was captured using the UK Biobank Axiom Array (Applied Biosystems) which contains 825,927 markers. The vitamin D GWAS study conducted a genome-wide association of 25-hydroxyvitamin D (25(OH)D) concentration in 417580 individuals from the UKB and combined them in a meta-analysis with results from a previous GWAS of 79,366 Euro-

peans (Revez et al., 2020; Jiang et al., 2018). 25(OH)D levels were measured using DiaSorin LIASON® chemiluminescent immunoassay which has a detection range of 10 nmol/L < 25(OH)D < 375 nmol/L (Revez et al., 2020).

To date, there is only one GWAS of serum 25(OH)D on individuals of Asian ancestry which recruited 3538 individuals from an Indian population in Punjabi (Sapkota et al., 2016). This study replicated an association between GC ($p = 0.007$) and CYP2R1 ($p = 0.019$) loci with 25(OH)D concentrations (T. J. Wang et al., 2010). However, due to the low number of study participants, I choose to use SNPs that were found to be associated with serum 25(OH)D levels in previous Asian MR studies (T. Huang et al., 2019; Lu et al., 2018). These MR studies genotyped 95,680 individuals randomly selected from the China Kadoorie Biobank (CKB) using a 384-SNP panel (Illumina GoldenGate). Specific SNPs (CYP2R1-rs10741657, DHCR7-rs12785878, GC-rs2282679, CYP24A1-rs6013897 identified from a previous genome-wide study in Europeans³² were included in the genotyping panel. SNPs in the DHCR7, CYP2R1, and GC loci were significantly associated with plasma 25(OH)D concentration while the CYP24A1 SNP had a consistent direction of effect with the European GWAS although the association was not significant (Lu et al., 2018).

3.1.2 TB GWAS Studies

The outcome data were summary statistics from eight individual tuberculosis GWAS studies across Asia and Europe from the International Tuberculosis Host Genetics Consortium (ITHGC) (Figure 5). This ongoing collaboration has collated a total of 15,573 TB cases and 24,843 controls (5309 cases and 8033 controls of Asian ancestry and 6739 cases and 13386 controls of European ancestry) genotyped data.

International Tuberculosis Host Genetics Consortium

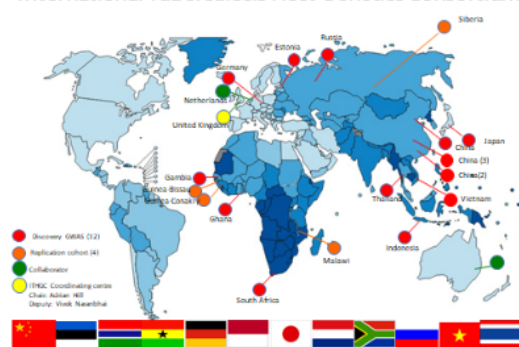


Figure 5. Map of the countries participating in the International Tuberculosis Host Genetics Consortium

3.2 MR Assumptions

Different assumptions have to be met while conducting a Mendelian randomization analysis. The first instrumental variable (IV) assumption is the assumption of relevance that states that the genetic variant should be associated with the serum-25(OH)D levels. The second IV assumption is the assumption of independence, which states that the variant should not be associated with any confounder of the serum-25(OH)D levels- TB association. The third IV assumption is the exclusion restriction, which states that the association between the variant and TB should only be through the serum-25(OH)D levels. These assumptions were addressed in the selection of genetic variants and also in the sensitivity analyses as described in the sections below.

3.3 Selection of genetic variants

The instrumental variables were single nucleotide polymorphisms (SNPs) associated with serum vitamin D levels (25(OH)D) at genome-wide significance ($p < 5 \times 10^{-8}$) (Table 3). I chose eight SNPs with the lowest p-values based on reported GWAS associations and their use in previous MR studies (Aspelund et al., 2019; Larsson et al., 2018; Mokry et al., 2015). These SNPs are on the GC (rs2282679), CYP24A1 (rs17216707 and rs6013897), DHCR7/NADSYN1 (rs11234027 and rs12785878) and CYP2R1 (rs12794714 and rs10741657) genes. The genes encode proteins in the vitamin D synthesis and metabolic pathway and since they have biological relevance to exposure, they are more plausible to the assumption of relevance. To ensure that the SNPs were independent, I used the 1000 genomes project data as a reference cluster to estimate linkage disequilibrium between the SNPs. SNPs that had a coefficient of determination ($R^2 = 1$) were in linkage disequilibrium and therefore the SNP with the lowest p-value in their association with serum-25(OH)D were retained.

I systematically screened the selected variants in PhenoScanner (<http://www.phenoscanter.medschl.cam.ac.uk/>), a database that gives the human genotype-phenotype association, to ensure the variants were not directly associated with any confounder of the exposure-outcome association in fulfilment of the MR assumption of independence. Further testing for the fulfilment of the instrumental variable assumptions was done in the sensitivity analyses discussed below.

I calculated the statistical power for the Mendelian randomization with a binary outcome using a web-based application (<https://sb452.shinyapps.io/power/>). The input variables included: the TB data sample size; the ratio of TB cases to controls; coefficient of determination (R^2) of serum vitamin D levels on genetic variants (13%), as reported in the vitamin D GWAS (Revez et al., 2020); the change in outcome per standard error change in exposure as reported in an observational study (2.57) (S. J. Huang et al., 2017); and a type-I error rate of 0.05.

3.4 Mendelian Randomization Analyses

All Mendelian randomization (MR) tests were done in the statistical software R version 3.6.1 (Team et al., 2020), using the TwoSampleMR R Package (<https://mrcieu.github.io/TwoSampleMR/authors.html>) (Hemani et al., 2018).

3.4.1 Variant Harmonization

I performed variant harmonization to ensure that the effect allele of the selected variant on the serum-25(OH)D dataset is the same effect allele on the TB dataset (Table 4).

3.4.2 Primary Analysis

In the primary analysis, I used the inverse-variance weighted (IVW) method. Mathematically, we can characterise an instrumental variable (IV) as a random variable by assuming that we have an outcome Y that is a function of a measured exposure X and unmeasured confounders U. We express X as a function of the genetic variant G and the confounder U. We, therefore, rewrite the IV assumptions as:

- I. G is associated with X
- II. G is independent of U
- III. G is independent of Y conditional on X and U

Let $\hat{\beta}_{Yj}$ be the genetic association with a standard error σ_{Yj} of an individual variant $j=1, \dots, 5$ with TB while $\hat{\beta}_{Xj}$ is the genetic association with a standard error σ_{Xj} of the variant with serum-25(OH)D concentrations. The genetic variants are not in linkage disequilibrium and are therefore assumed to be independently distributed. The genetic variants are also oriented such that the $\hat{\beta}_{Xj}$ estimates are all either positive or negative. The IVW estimate is calculated by a meta-analysis of each variant's ratio estimate. The ratio estimate of the j^{th} genetic variant is denoted as $\hat{\beta}_{IVj}$ and is calculated as: (Lawlor, Harbord, Sterne, Timpson, & Davey Smith, 2008)

$$\hat{\beta}_{IVj} = \frac{\hat{\beta}_{Yj}}{\hat{\beta}_{Xj}} \quad (1)$$

Since I was using non-overlapping datasets, I assumed that the correlation between $\hat{\beta}_{Yj}$ and $\hat{\beta}_{Xj}$ is zero (Pierce & Burgess, 2013), hence variance is:

$$\text{Var}(\hat{\beta}_{IVj}) = \frac{\sigma_{Yj}^2}{\hat{\beta}_{Xj}^2} \quad (2)$$

and the standard error is:

$$SE(\hat{\beta}_{IVj}) = \frac{\sigma_{Yj}}{\hat{\beta}_{Xj}} \quad (3)$$

¹³ The IVW estimate is a weighted mean of the individual variant causal estimates which is calculated as: (Borenstein, Hedges, Higgins, & Rothstein 2011)

$$\hat{\beta}_{IVW} = \frac{\sum_j W_j \hat{\beta}_{IVj}}{\sum_j W_j} \quad (4)$$

where

$$W_j = SE(\hat{\beta}_{IVj})^{-2} \quad (5)$$

are the inverse variance weights. The standard error (assuming a fixed effect model) is:

$$SE(\hat{\beta}_{IVW}) = \sqrt{\frac{1}{\sum_j \hat{\beta}_{Xj}^2 \sigma_{Yj}^{-2}}} \quad (6)$$

I used the multiplicative random-effect IVW model. This model ensures that the relative weighting of each variant estimate does not change. The model also calculates the heterogeneity explained by each variant's causal estimate (Bowden et al. 2017). This model assumes that the $\hat{\beta}_{IVj}$ estimates are normally distributed with mean β and variance $\phi_M^2 \sigma_{IVj}^2$.

$$\hat{\beta}_{IVj} \sim N(\beta, \phi_M^2 \sigma_{IVj}^2) \quad (7)$$

¹³ This model allows the estimation of the residual standard error, which is equivalent to the heterogeneity parameter ϕ_M as part of the model. The point estimate from a multiplicative random-effects model is similar to the point estimate from a fixed-effect model (Thompson & Sharp 1999). This model also assumes balanced pleiotropy (pleiotropic effects on TB has an equal possibility of either being positive or negative) to provide valid causal estimates (Bowden et al. 2017). I went ahead and performed sensitivity analyses to test how robust of our primary analysis results are. Analyses were done separately for each country and then pooled for each broad ethnicity.

3.4.3 Sensitivity Analysis

The robust analyses are meant to address horizontal pleiotropy (the MR assumption of exclusion restriction). They also allow the inference of valid causal effect although their assumptions are weaker compared to the primary analysis assumptions (Burgess et al. 2019). I used the weighted median regression and MR-Egger analyses for the sensitivity

analysis²⁵ MR-Egger analysis is a weighted linear regression model of the $\hat{\beta}_{Yj}$ on the $\hat{\beta}_{Xj}$ where all the $\hat{\beta}_{Xj}$ associations are oriented to be all either positive or negative (Bowden, Davey Smith, & Burgess, 2015).

$$\hat{\beta}_{Yj} = \alpha_{0E} + \alpha_E \hat{\beta}_{Xj} \quad (8)$$

The intercept α_{0E} estimates the average horizontal pleiotropic effect (direct effect of the variants on TB that are not through serum-25(OH)D levels). The causal effect is the slope α_E estimated from a weighted regression of the genetic variant-TB association on the genetic variant-serum 25(OH)D levels association. The model is similar to that of IVW except that it includes an intercept term (α_E equals $\hat{\beta}_{IVW}$ when α_{0E} is zero.) When the intercept differs from zero, then there is horizontal pleiotropy and hence the IVW estimator is biased. The α_E estimate represents the true effect of causality Under the Instrument Strength Independent of Direct Effect (InSIDE) assumption. (Bowden et al., 2015)²⁰ This assumption allows genetic variants to have pleiotropy independent of the variant-serum 25(OH)D levels associations and hence some heterogeneity in the causal estimates of individual variants may be observed. This heterogeneity does not invalidate the analysis since it is a consequence of weakening IV assumptions. The weighted median regression works under the majority valid assumption which states that 50% of the selected genetic variants are valid IVs (Bowden et al., 2015)²⁵. The weighted median estimator is the median of a distribution having $\hat{\beta}_{IVj}$ as its

$$P_{IVj} = 100(S_j - \frac{W_j}{2}) \quad (9)$$

th percentile, where

$$S_j = \sum_{j=1}^j W_j \quad (10)$$

This method takes the variant-specific causal estimate and calculates the central tendency of the select estimates. The median of the distribution is not influenced by the outlying estimates.

3.4.4 Scatter Plot and Test for Heterogeneity

Using the Cochran's Q test, I did a heterogeneity test to establish whether the IV causal estimates vary based on the individual genetic variant (Bowden et al., 2019)²². This test relies on the assumption that all the genetic variants identify a similar causal parameter. H_0 : IV causal estimates are homogeneous H_1 : IV causal estimates are heterogeneous The statistic is calculated as:

$$Q = \sum_j W_j (\hat{\beta}_{IVj} - \hat{\beta}_{IVW})^2 \quad (11)$$

and has a chi-square distribution with $J-1$ degrees of freedom, where, J denotes the total genetic variants. Besides, I did an I-squared (I^2) heterogeneity test to measure the percentage of variability in the effect estimates. This is calculated as:

$$I^2 = \left(\frac{Q - df}{Q}\right) \times 100\% \quad (12)$$

Where Q is the Cochran's Q statistic and df is the degree of freedom of the Q . This was also visualised using a scatter plot of the genetic association with TB against the genetic association with serum-25(OH)D levels, including their confidence intervals. Each point on the graph represents an individual genetic variant. Under the null hypothesis, these points should lie on a straight line through the origin. Variants that lay outside the line were tested for pleiotropy.

3.4.5 Funnel Plot and Test for Directional Pleiotropy

In addition to the MR-Egger test for directional pleiotropy, I also used funnel plots. A funnel plot is used to measure the precision of the IV estimates and to test for directional pleiotropy (Bowden et al. 2015). I plotted the reciprocal of the standard error of the IV estimates

$$\frac{1}{SE(\hat{\beta}_{IVj})} = \frac{\hat{\beta}_{Xj}}{\sigma_{Yj}} \quad (13)$$

against the IV estimates

$$\frac{\hat{\beta}_{Yj}}{\hat{\beta}_{Xj}} \quad (14)$$

Asymmetry in the funnel plot signifies horizontal pleiotropy. This indicates that the estimated causal effect of the risk factor on the outcome may be biased.

3.5 Meta-analyses

I pooled the country IVW estimates to represent two ethnic populations; Asian and European and conducted meta-analyses. I also pooled the IVW estimates for all the countries and carried out an overall meta-analysis. I used the I^2 heterogeneity test to measure the percentage of variability of the effect estimates. I also used the tau-squared (τ^2) which explains the between-study (between-country in this case) variance. I used both the random and fixed effect models to calculate the pooled effect estimate of serum 25(OH)D levels on tuberculosis. I used the metagen function in the meta package of R statistical software (Balduzzi, Rücker, & Schwarzer 2019).

4 Chapter 4: Results

4.0.1 Extraction of the Instrumental Variables

I conducted Mendelian randomization (MR) analyses to determine the effect of genetically predicted serum 25-hydroxyvitamin D (25(OH)D) on the odds of tuberculosis disease. Instrumental variables (IV) were extracted from the UK Biobank vitamin D G³²AS and were genetic variants (SNPs) that had a genome-wide association ($p = 510^{-8}$) with serum-25(OH)D77 (Table 3). SNPs on the same loci were found to be in linkage disequilibrium ($R^2 = 1$) after clumping to test for independence of the IVs. The SNP with the lowest p-value in their association with serum-25(OH)D were retained and I, therefore, remained with four IVs. The effect allele of all the SNPs were oriented such that the effect size estimates (beta) of the genetic association with serum-25(OH)D were all positive. I³⁴ extracted the four IVs from the different outcome datasets. I did harmonization to ensure that the effect allele of an IV on the exposure dataset was the same effect allele on the outcome dataset (Table 4-5).

Table 1. Effects of genetic variants on serum 25(OH)D levels

SNP	Gene	EA/OA	EAF	N	Beta	SE	P-value	Var
rs10741657	CYP2R1	A/G	0.40	417580	0.09	0.002	$1.00e^{-300}$	0.0038
rs12785878	DHCR7/NADSYN1	T/G	0.79	417580	0.11	0.002	$1.00e^{-300}$	0.0040
rs2282679	GC	T/G	0.71	417580	0.19	0.002	$1.00e^{-300}$	0.0150
rs6013897	CYP24A1	T/A	0.80	417041	0.04	0.003	$4.25e^{-50}$	0.0005

EA, effect allele; OA, other allele; EAF, effect allele frequency; N, sample size; Beta, effect size of the serum-25(OH)D-increasing allele on rank-based inverse normal transformed (RINT) 25OHD levels; SE, standard error; Var, variance in 25(OH)D levels explained by each SNP

China1 data						
SNP	N	EA/OA	EAF	Beta	SE	P-value
rs1074165	1070	A/G	0.38	0.04	0.13	0.37
rs12785878	1070	T/G	0.48	-0.04	0.13	0.35
rs2282679	1070	T/G	0.67	0.06	0.13	0.33
rs6013897	1070	T/A	0.83	0.06	0.17	0.37
China2 data						
SNP	N	EA/OA	EAF	Beta	SE	P-value
rs10741657	2390	A/G	0.38	0.01	0.06	0.41

rs12785878	2390	T/G	0.52	-0.06	0.06	0.17
rs2282679	2390	T/G	0.69	0.13	0.06	0.02
rs6013897	2390	T/A	0.84	0.10	0.08	0.12
China3 data						
SNP	N	EA/OA	EAF	Beta	⁷ SE	P-value
rs10741657	2509	A/G	0.37	-0.15	0.06	0.01
rs12785878	2509	T/G	0.48	0.02	0.06	0.34
rs2282679	2509	T/G	0.69	0.09	0.06	0.07
rs6013897	2509	T/A	0.84	-0.02	0.08	0.37
Japan data						
SNP	N	EA/OA	EAF	Beta	⁷ SE	P-value
rs10741657	3950	A/G	0.36	-0.05	0.06	0.21
rs12785878	3950	T/G	0.33	-0.02	0.06	0.38
rs2282679	3950	T/G	0.73	-0.02	0.07	0.36
rs6013897	3950	T/A	0.91	0.10	0.12	0.18
Thailand data						
SNP	N	EA/OA	EAF	Beta	⁷ SE	P-value
rs10741657	1431	A/G	0.3	0.05	0.08	0.27
rs12785878	1431	T/G	0.3	-0.1	0.09	0.14
rs2282679	1431	T/G	0.75	0.15	0.12	0.1
rs6013897	1431	T/A	0.81	-0.11	0.1	0.15
Estonia data						
SNP	N	EA/OA	EAF	Beta	⁷ SE	P-value
rs10741657	7286	A/G	0.43	0.05	0.10	0.29
rs12785878	7286	T/G	0.65	0.18	0.10	0.03
rs2282679	7286	T/G	0.72	-0.03	0.10	0.38
rs6013897	7285	T/A	0.82	0.21	0.12	0.04
Germany data						
SNP	N	EA/OA	EAF	Beta	⁷ SE	P-value
rs10741657	919	A/G	0.39	0.09	0.11	0.21

rs12785878	919	T/G	0.68	0.04	0.11	0.37
rs2282679	919	T/G	0.73	0.04	0.12	0.37
rs6013897	919	T/A	0.81	-0.15	0.14	0.13
Russia data						
SNP	N	EA/OA	EAF	Beta	SE	P-value
rs10741657	11445	A/G	0.42	-0.03	0.03	0.13
rs12785878	11445	T/G	0.68	-0.05	0.03	0.04
rs2282679	11445	T/G	0.70	0.04	0.03	0.09
rs6013897	11445	T/A	0.83	-0.002	0.04	0.47

Table 2. Effect of the instrumental variables on the odds of tuberculosis disease in each study country

4.1 Mendelian Randomization Results

I had 100% statistical power in all the countries to detect a causal association between serum-25(OH)D concentrations and the odds of tuberculosis disease except in Estonia which had 99.8% power and Germany with 99.9% power. The primary estimates, which were calculated by the inverse variance weighted (IVW) method revealed a lack of causal association between genetically elevated serum-25(OH)D and the risk of TB disease ($p > 0.05$) (Table 6-7). The sensitivity analyses using MR-Egger and weighted median confirmed the lack of association (Table 8). These results were also visualised using forest plots (Figure 6).

China1 results				
SNP	Sample size	Beta	SE	P-value
rs10741657	1070	0.48	1.43	0.73
rs12785878	1070	-0.44	1.15	0.70
rs2282679	1070	0.30	0.70	0.67
rs6013897	1070	1.54	4.48	0.73
All - Inverse variance weighted	1070	0.18	0.55	0.74
All - MR Egger	1070	0.08	1.31	0.96
All - Weighted median	1070	0.24	0.57	0.70
China2 results				
SNP	Sample size	Beta	SE	P-value

rs10741657	2390	0.16	0.69	0.81
rs12785878	2390	-0.52	0.55	0.34
rs2282679	2390	0.67	0.33	0.04
rs6013897	2390	2.62	2.18	0.23
All - Inverse variance weighted	2390	0.35	0.33	0.28
All - MR Egger	2390	0.51	0.95	0.65
All - Weighted median	2390	0.43	0.30	0.15
China3 results				
SNP	Sample size	Beta	SE	P-value
rs10741657	2509	-1.66	0.69	0.02
rs12785878	2509	0.22	0.54	0.69
rs2282679	2509	0.49	0.34	0.15
rs6013897	2509	-0.74	2.18	0.73
All - Inverse variance weighted	2509	0.10	0.43	0.82
All - MR Egger	2509	1.17	0.95	0.34
All - Weighted median	2509	0.36	0.30	0.23
Japan results				
SNP	Sample size	Beta	SE	P-value
rs10741657	3950	-0.54	0.68	0.43
rs12785878	3950	-0.17	0.58	0.77
rs2282679	3950	-0.12	0.34	0.73
rs6013897	3950	2.60	2.86	0.37
All - Inverse variance weighted	3950	-0.17	0.27	0.53
All - MR Egger	3950	-0.33	0.70	0.68
All - Weighted median	3950	-0.14	0.29	0.62
Thailand results				
SNP	Sample size	Beta	SE	P-value
rs10741657	1431	0.56	0.93	0.55
rs12785878	1431	-0.88	0.80	0.27
rs2282679	1431	0.78	0.61	0.20

rs6013897	1431	-2.82	2.69	0.29
All - Inverse variance weighted	1431	0.18	0.50	0.72
All - MR Egger	1431	1.43	1.02	0.30
All - Weighted median	1431	0.62	0.50	0.22
Estonia results				
SNP	Sample size	Beta	SE	P-value
rs10741657	7286	0.58	1.05	0.58
rs12785878	7286	1.66	0.90	0.07
rs2282679	7286	-0.16	0.54	0.76
rs6013897	7286	5.62	3.26	0.08
All - Inverse variance weighted	7286	0.45	0.58	0.44
All - MR Egger	7286	-1.41	0.99	0.29
All - Weighted median	7286	0.22	0.48	0.65
Germany results				
SNP	Sample size	Beta	SE	P-value
rs10741657	919	0.98	1.23	0.42
rs12785878	919	0.34	1.02	0.74
rs2282679	919	0.20	0.61	0.74
rs6013897	919	-4.04	3.60	0.26
All - Inverse variance weighted	919	0.27	0.48	0.57
All - MR Egger	919	0.85	1.11	0.52
All - Weighted median	919	0.26	0.51	0.62
Russia results				
SNP	Sample size	Beta	SE	P-value
rs10741657	11445	-0.35	0.31	0.26
rs12785878	11445	-0.47	0.28	0.09
rs2282679	11445	0.22	0.16	0.18
rs6013897	11445	-0.07	1.02	0.95
All - Inverse variance weighted	11445	-0.02	0.18	0.90
All - MR Egger	11445	0.42	0.39	0.40

All - Weighted median	11445	0.04	0.15	0.77
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Table 3. Mendelian randomization results for the effect of genetically elevated serum-25(OH)D₂ on the odds of tuberculosis disease

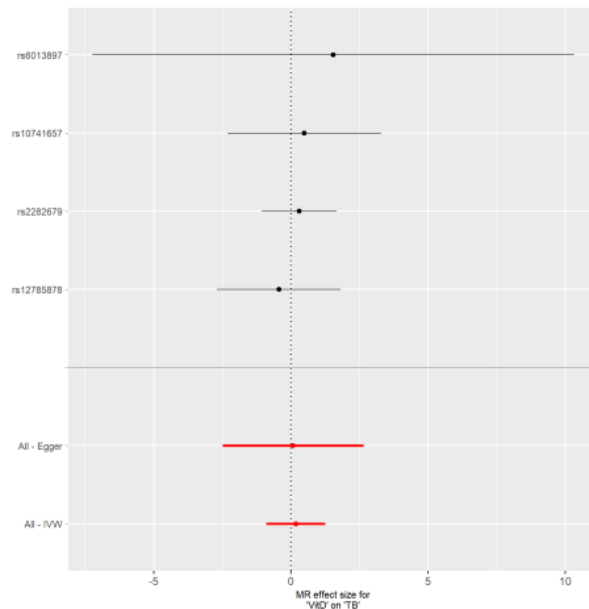


Figure 6. China 1

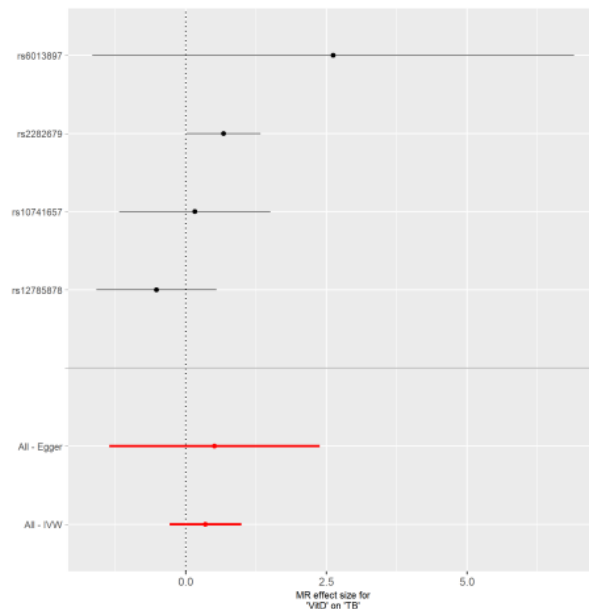


Figure 7. China 2

Figure 8. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

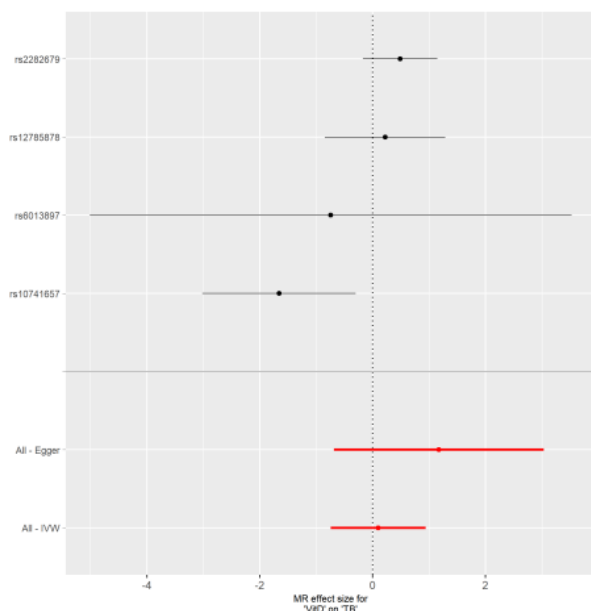


Figure 8. China 3

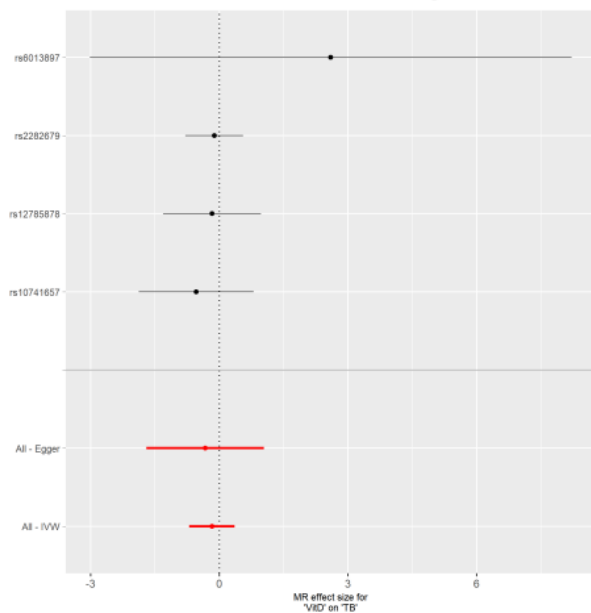


Figure 9. Japan

Figure 10. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

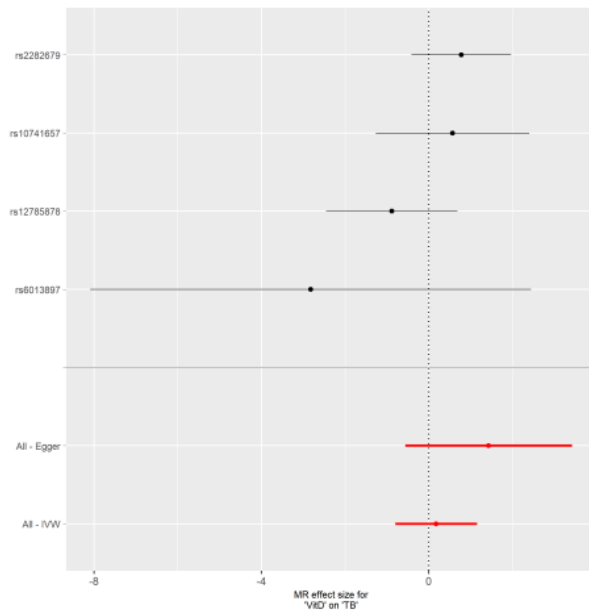


Figure 10. Thailand

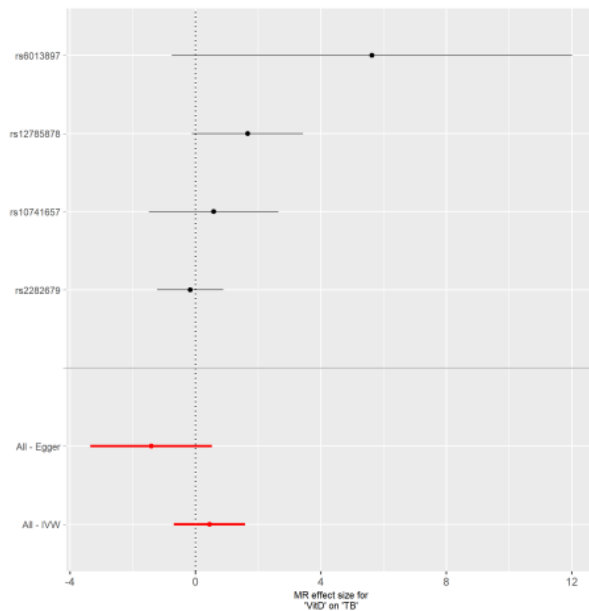


Figure 11. Estonia

Figure 12. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

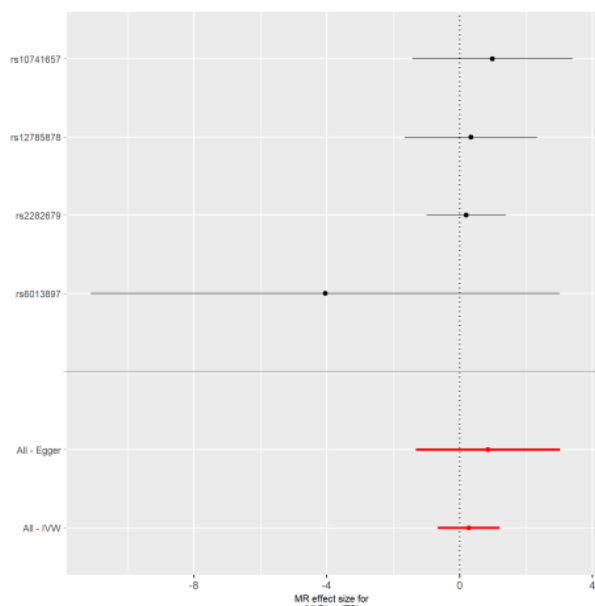


Figure 12. Germany

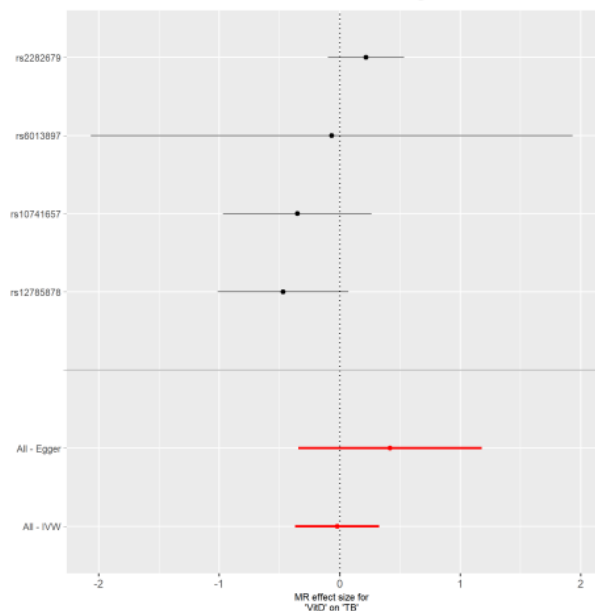


Figure 13. Russia

Figure 14. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

4.1.1 Heterogeneity and Pleiotropy Results

Heterogeneity was initially assessed using a scatter plot of genetic association with tuberculosis against the genetic association with serum-25(OH)D concentrations (Figure 7). Most of the genetic variants (points on the graph) in the different countries lied away

from the straight line through the origin and were therefore tested for pleiotropy. Further tests for heterogeneity of the IV estimates based on individual genetic variant were done using the Cochran's Q test under the null hypothesis of homogeneity. Results for the Cochran's Q test showed a lack of heterogeneity ($P \geq 0.05$) among the genetic variants and therefore confirms that our select IVs were valid (Table 6). The amount of heterogeneity was expressed using the I-squared statistic. An I-squared below 50% represents homogeneity and only one of the country had slight heterogeneity (China 3; $I^2 = 62\%$) (Table 9). The MR-Egger intercepts are the average pleiotropic effect of a genetic variant. Under the null hypothesis of no pleiotropy, the intercepts should not differ from zero. Results of the test for directional pleiotropy showed no evidence of pleiotropy ($P \geq 0.05$) (Table 9). These results further validated our choice of genetic variants.

Table 4. Heterogeneity and Pleiotropy results

Country	Q	Q-df	Q-pval	I^2	egger-intercept	egger-se	egger-pval
China1	0.46	3	0.93	0	0.01	0.16	0.94
China2	4.64	3	0.20	35	-0.02	0.12	0.87
China3	7.97	3	0.05	62	-0.15	0.12	0.34
Japan	1.25	3	0.74	0	0.02	0.09	0.83
Thailand	4.14	3	0.25	28	-0.16	0.11	0.31
Estonia	5.65	3	0.13	47	0.25	0.12	0.17
Germany	1.79	3	0.62	0	-0.08	0.14	0.62
Russia	5.96	3	0.11	50	-0.06	0.05	0.34

Cochran's Q statistic for heterogeneity; Qdf, Q statistic degrees of freedom; I^2 , test for heterogeneity; Egger, intercept, MR – Egger measure of pleiotropy; Egger, se, MR – Egger standard error; Egger, p, val, MR – Egger p value

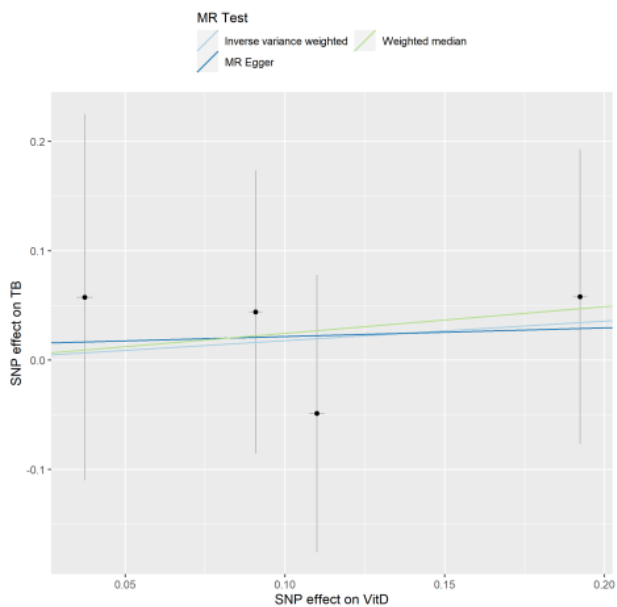


Figure 15. China 1

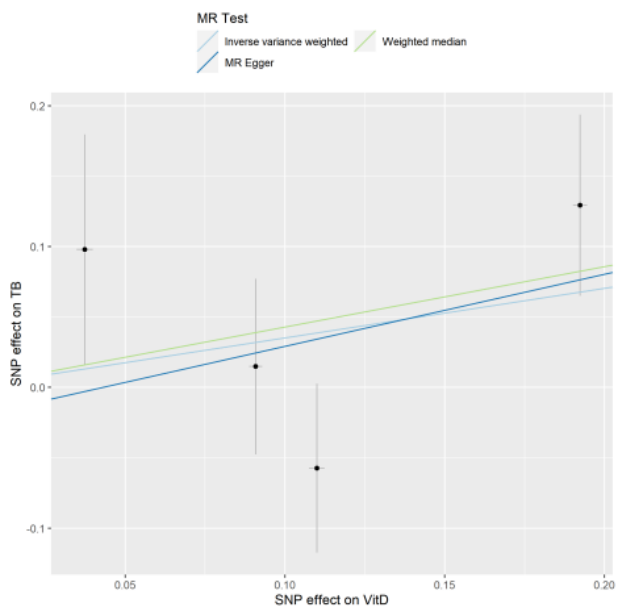


Figure 16. China 2

Figure 17. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

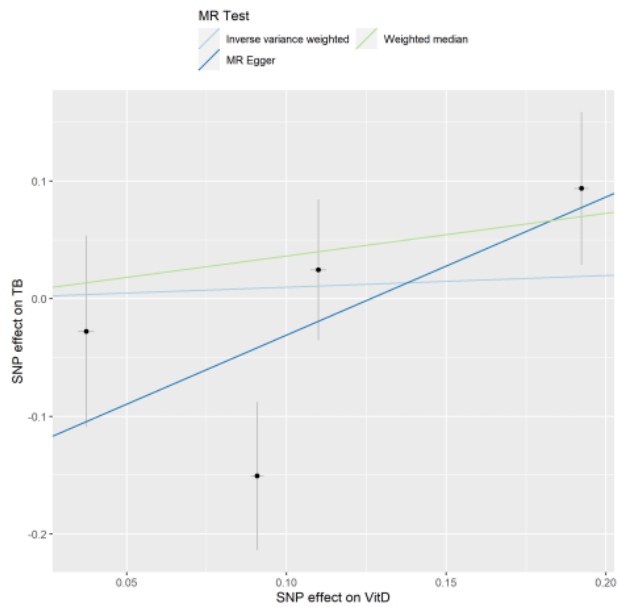


Figure 17. China 3

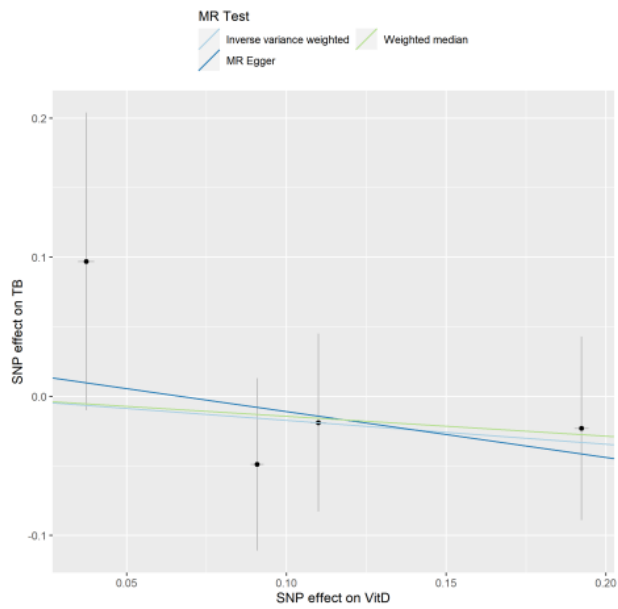


Figure 18. Japan

26
Figure 19. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

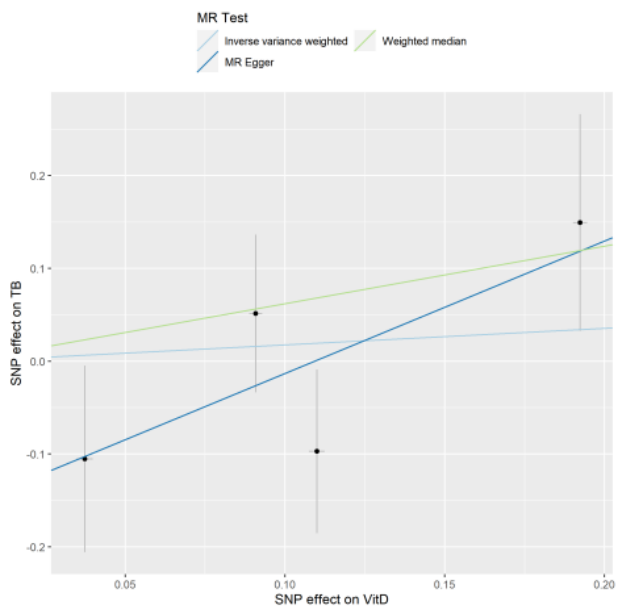


Figure 19. Thailand

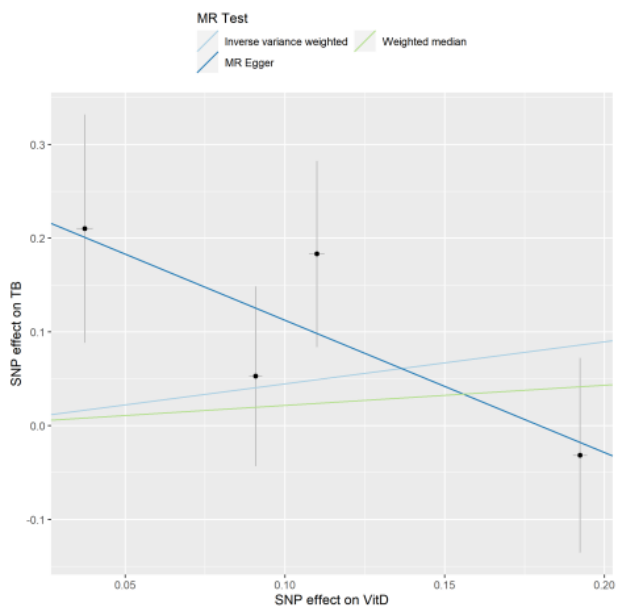


Figure 20. Estonia

Figure 21. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

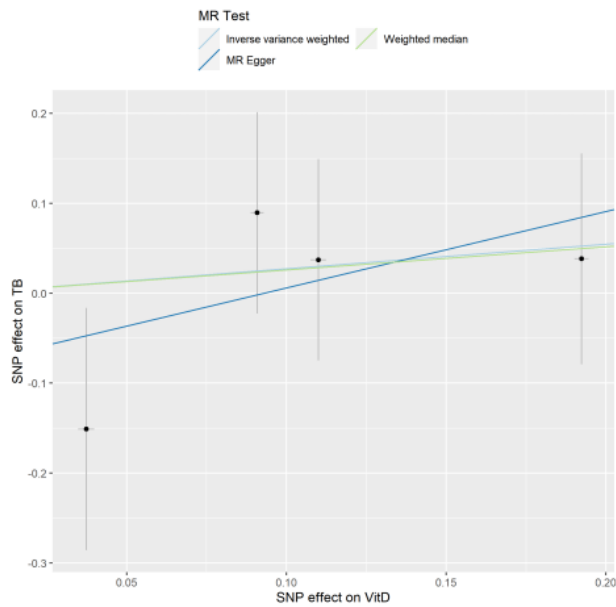


Figure 21. Germany

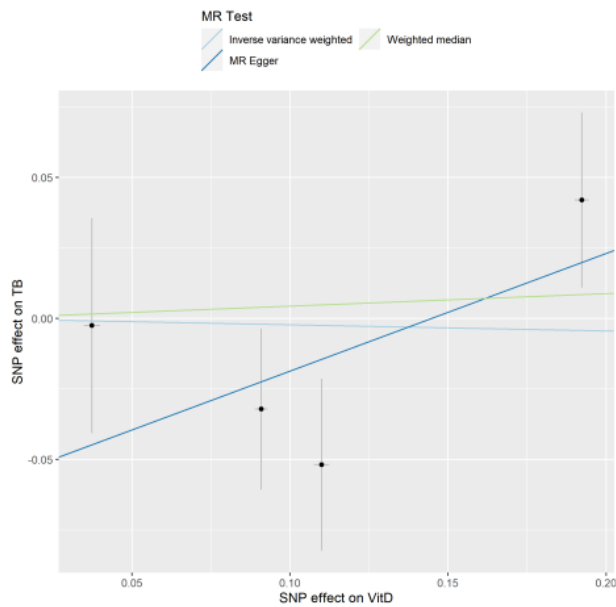


Figure 22. Russia

Figure 23. Forest plots results for the effect of genetically predicted serum-25(OH)D on the odds of tuberculosis disease

4.2 Meta-analyses

Using the Mendelian randomization inverse variance weighted (MR-IVW) results, I performed meta-analyses using the inverse variance weighted (IVW) - fixed effect model to test the overall association between genetically elevated serum-25(OH)D levels and the

odds of tuberculosis disease. I also tested the association within the Asian population and European population by doing meta-analyses of the MR-IVW estimates of the respective countries.

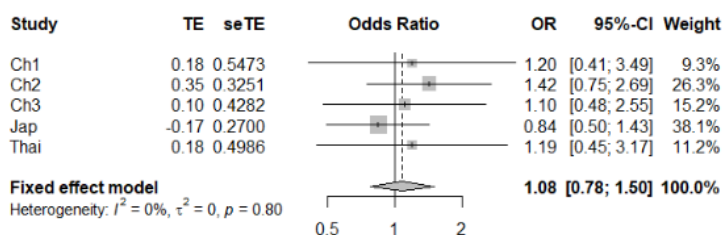


Figure 24. Meta-analysis of the MR-IVW estimates of the Asian countries

TE, Treatment effect which is the MR-IVW estimate of the odds of tuberculosis disease per standard deviation increase in genetically determined serum-25(OH)D levels; seTE, standard error of the treatment effect; OR, Odds ratio, 95%-CI, 95% confidence interval; I^2 , I-squared test for heterogeneity; τ^2 , tau-squared test for heterogeneity; p, p value; Ch1, China1 data; Ch2, China 2 data; Ch3, China 3 data; Jap, Japan data; Thai, Thailand data

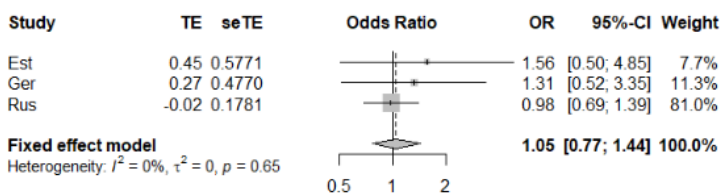


Figure 25. Meta-analysis of the MR-IVW estimates of the European countries

TE, Treatment effect which is the MR-IVW estimate of the odds of tuberculosis disease per standard deviation increase in genetically determined serum-25(OH)D levels; seTE, standard error of the treatment effect; OR, Odds ratio, 95%-CI, 95% confidence interval; I^2 , I-squared test for heterogeneity; τ^2 , tau-squared test for heterogeneity; p, p value; Est, Estonia data; Ger, Germany data; Rus, Russia data

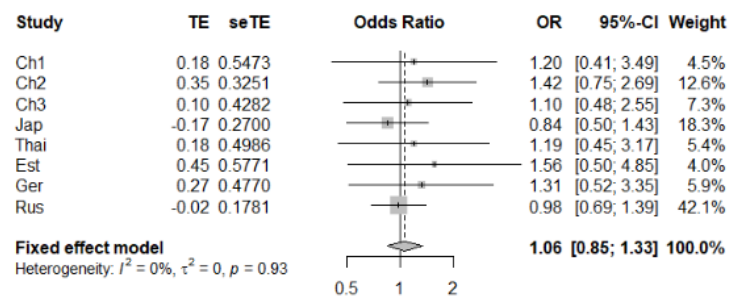


Figure 26. Meta-analysis of the MR-IVW estimates of both Asian and European countries

TE, Treatment effect which is the MR-IVW estimate of the odds of tuberculosis disease per standard deviation increase in genetically determined serum-25(OH)D levels; seTE, standard error of the treatment effect; OR, Odds ratio, 95%-CI, 95% confidence interval; I^2 , I-squared test for heterogeneity; τ^2 , tau-squared test for heterogeneity; p, p value; Ch1, China1 data; Ch2, China 2 data; Ch3, China 3 data; Jap, Japan data; Thai, Thailand data; Est, Estonia data; Ger, Germany data; Rus, Russia data

Meta-analyses of the IVW estimates showed no association between genetically elevated serum 25(OH)D levels and the odds of TB overall (odds ratio [OR]=1.08, 95% confidence interval [CI]=0.85-1.33, $p=0.93$) or in the two ethnic populations, European (OR=1.05, 95% CI=0.77-1.44, $p=0.65$) and Asian (OR=1.08, 95% CI=0.78-1.50, $p=0.80$) (Figure 8-10). I used the fixed-effect model within the meta-analysis since the IVW estimates had no evidence of both between and within-study heterogeneity ($I^2 = 0\%$, $\tau^2 = 0$).

5 Chapter 5: Discussion

Finding from our study suggest that genetically elevated serum-25(OH)D concentration were not causally associated with the odds of tuberculosis disease. The results are consistent with results from recently published randomized controlled trials (RCTs) testing the effect of vitamin D supplementation on the incidence of tuberculosis disease/infection. Sudfeld and colleagues found no impact of Vitamin D supplementation on mortality or the incidence of pulmonary tuberculosis in a large randomized controlled trial (RCT) enrolling adults living with HIV in Tanzania (Sudfeld et al., 2020). Vitamin D supplementation also showed no effect in the prevention of tuberculosis infection and disease in an RCT that enrolled children in Mongolia (Ganmaa et al., 2020). The difference in our study compared to the RCTs was the potential to capture all the sources of vitamin D which includes exposure to sunlight, diet and supplementation by use of genetic variants that affect serum-25(OH)D rather than supplementation alone as in RCTs. Besides, lifelong exposure to vitamin D is tested rather than the time-limited supplementation. Nonetheless, RCTs remain the gold standard and are the most robust means of evaluating potential interventions. The strengths of this study include the lack of potential confounding by environmental factors. These could be a possible explanation of the reported association between low vitamin D and the risk of TB reported in many observational studies. Since the outcome of disease does not affect the genetic make-up of an individual, the study also eliminated possible effects of reverse causation which might be found in observational studies. The study also had enough power to detect the causal association between serum-25(OH)D levels and TB disease. In addition, there were no violations of the assumptions of Mendelian randomization (MR) as far as they could be tested. Potential limitations of this study just like any other MR study is that I could not fully rule out the effect of possible genetic pleiotropy and linkage disequilibrium (LD) between the genetic variants that I used as instrumental variables. However, findings from the pleiotropy and independence tests showed no evidence of horizontal pleiotropy or LD to support the possibility. The cohorts in this study included individual of European and Asian ancestry and therefore these results may not apply to other ethnic populations. In conclusion, genetically elevated serum-25(OH)D does not has a causal association with tuberculosis disease.

5.1 Future Research

I hope to study the effect of genetically determined serum-25(OH)D on the risk of tuberculosis in the African population in the near future.

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